

Model answer for T.D. no. 3 (Protein–ligand interactions)

Exercise n° 1 :

We have:

- ✓ The protein : glyceraldehyde-3-phosphate dehydrogenase with $[P_t] = 1\mu\text{M}$
- ✓ The ligand : NAD^+
- ✓ Technique used : tryptophan residue fluorescence

$[\text{NAD}^+]_{\text{total}} = [\text{NAD}^+]_{\text{bound}} + [\text{NAD}^+]_{\text{free}}$ so : $[\text{NAD}^+]_{\text{free}} = [\text{NAD}^+]_{\text{total}} - [\text{NAD}^+]_{\text{bound}}$

The graphical representation of KLOTZ consists of plotting $1/[\text{RL}]$ as a function of $1/[\text{L}]$, thus :

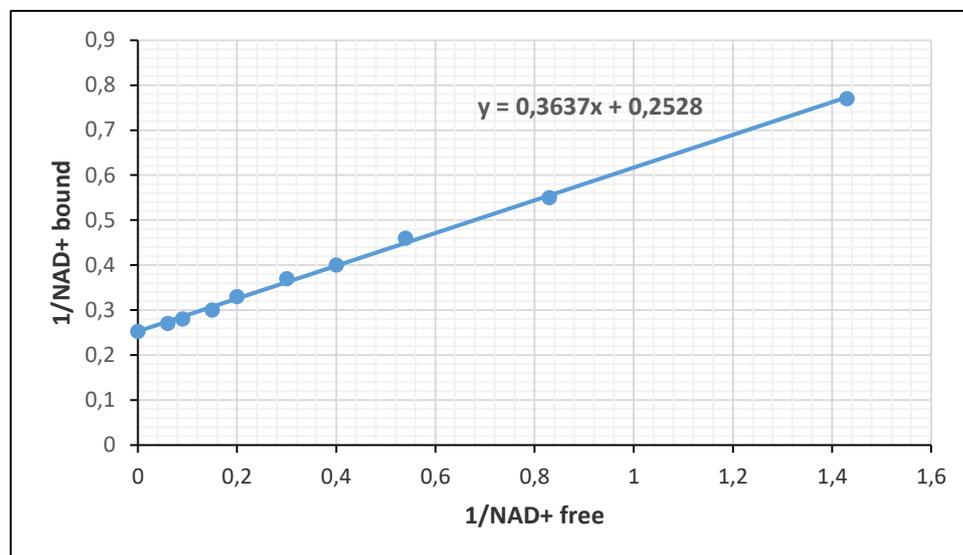
$$1/[\text{NAD}^+]_{\text{bound}} = f(1/[\text{NAD}^+]_{\text{free}})$$

$[\text{NAD}^+]_{\text{free}} (\mu\text{M})$	16,3	11,5	6,7	4,95	3,3	2,5	1,85	1,2	0,7
$1/[\text{NAD}^+]_{\text{bound}} (\mu\text{M}^{-1})$	0,27	0,28	0,30	0,33	0,37	0,40	0,46	0,55	0,77
$1/[\text{NAD}^+]_{\text{free}} (\mu\text{M}^{-1})$	0,06	0,09	0,15	0,20	0,30	0,40	0,54	0,83	1,43

The KLOTZ equation : $1/[\text{RL}] = (1/[\text{L}]) \cdot (K_D/n[P_t]) + 1/n[P_t]$

So: the slope = $K_D/n[P_t]$

y-intercept = $1/n[P_t]$



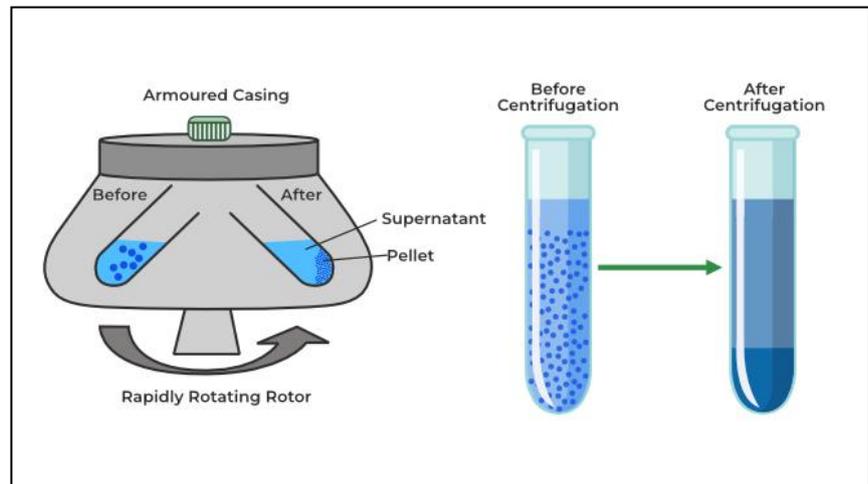
Graphically :

- ✓ y-intercept = $0,25 \mu\text{M}^{-1} = 1/n[P_t]$ so : $n = 4$ binding sites
- ✓ The slope = $0,396 = K_D/n[P_t]$ so : $K_D = 1,58 \mu\text{M}$
- ✓ $K_A = 1/K_D$ so : $K_A = 0,63 \mu\text{M}^{-1}$

Exercise n°2 :

We have :

- ✓ The protein : Na⁺/K⁺-dependent ATPase
- ✓ The ligand : rubidium ion (Rb⁺)
- ✓ The technique used : centrifugation

The experiment design :**The maximum amount of Rb⁺ bound to membranes (nmol/mg) :**

KLOTZ's graphical representation consists of plotting $1/[RL]$ as a function of $1/[L]$ so :

$$1/[Rb^+] \text{ bound} = f(1/[Rb^+] \text{ free})$$

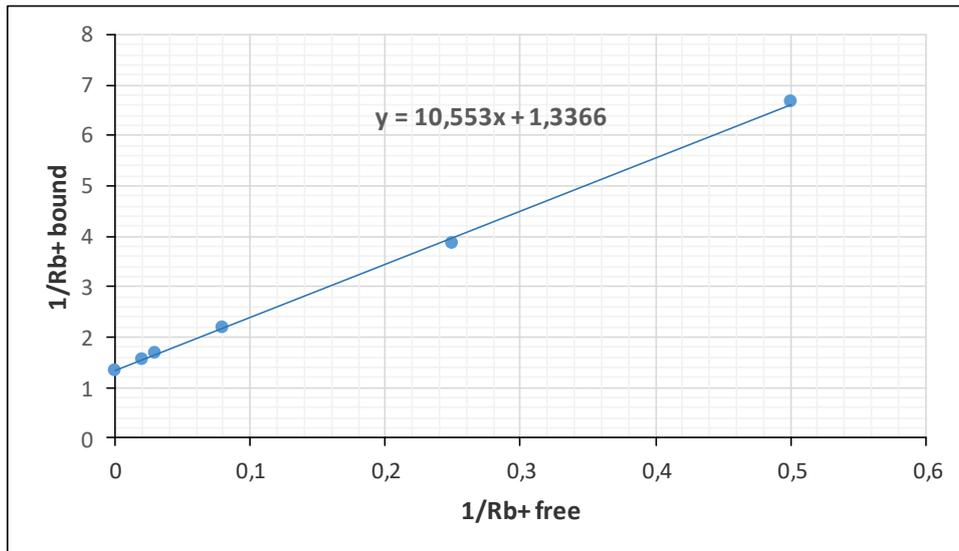
Knowing that : $[Rb^+]_{\text{in the supernatant}} = [Rb^+]_{\text{free}}$

$1/[Rb^+] \text{ free } (\mu\text{M}^{-1})$	0,50	0,25	0,08	0,03	0,02
$1/Rb^+ \text{ bound } (\text{nmol}^{-1})$	6,67	3,85	2,20	1,69	1,56

The **KLOTZ** equation : $1/[RL] = (1/[L]).(K_D/n[P_t]) + 1/n[P_t]$

So : the slope = $K_D/n[P_t]$

y-intercept = $1/n[P_t]$



Graphically :

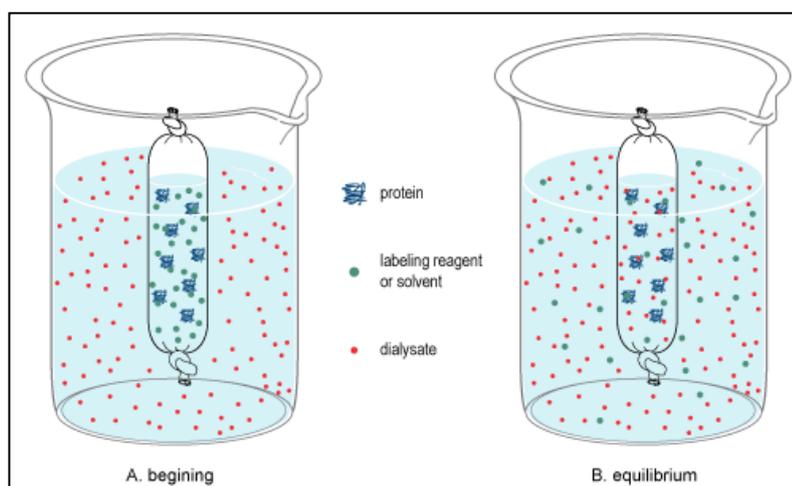
- y-intercept = $1,35 \text{ nmol}^{-1} = 1/n[P_t]$
- We have $[P_t] = 60 \mu\text{g} = 0,06\text{mg}$
- so $n = 1/1,35 \times 0,06$ from which **$n = 12,3 \text{ nmol/mg protein}$**
- The slope = $10,5 \text{ nmol}^{-1} \cdot \mu\text{M} = K_D/n[P_t]$ so : **$K_D = 7,75 \mu\text{M}$**
- We know that $K_A = 1/K_D$ so : **$K_A = 0,13 \mu\text{M}^{-1}$**

Exercise n° 3 :

We have :

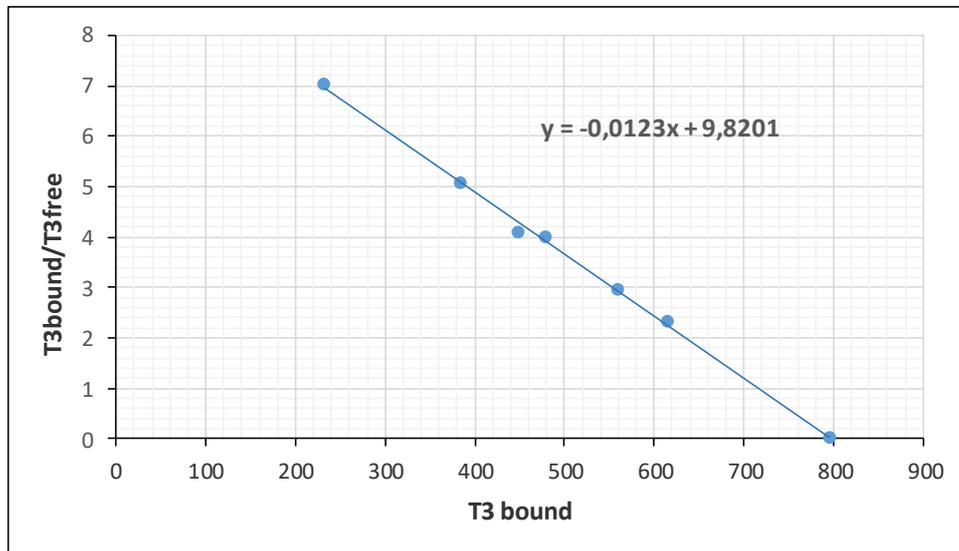
- ✓ $[T3]$ outside the dialysis bag = **$[T3]$ free**
- ✓ $[T3]$ inside the dialysis bag = **$[T3]$ bound + $[T3]$ free**

So : **$[T3]$ bound** = $[T3]$ inside the dialysis bag - $[T3]$ outside the dialysis bag



According to the SCATCHARD method : $[RL] / [L] = f([RL])$

[RL] = [T3] bound (nM)	232	384	450	480	560	615
[RL]/[L] = [T3] bound/[T3] free	7,03	5,05	4,09	4	2,95	2,32



Graphically :

- X-intercept : $n [Pt] = 810\text{nM}$
Knowing that : $[Pt] = 0,77\mu\text{M} = 770\text{nM}$ so : **$n = 1$ site**
- The slope: $-K_A = -0,012 \text{ nM}^{-1}$ so : **$K_A = 0,012 \text{ nM}^{-1}$**

Exercise n° 4 :

We have :

- The protein : calcium channels
- The ligand : verapamil
- The technique used : filtration and radioactivity

The graphical representation of **KLOTZ** consists of plotting $1/[RL]$ as a function of $1/[L]$, so :

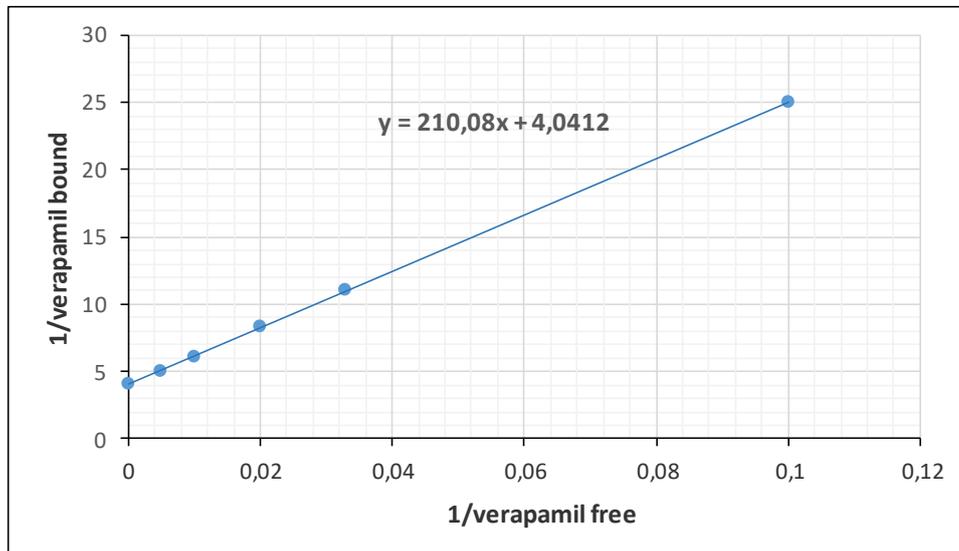
$$1/[\text{verapamil}] \text{ bound} = f(1/[\text{verapamil}] \text{ free})$$

$1/[\text{verapamil}] \text{ free (nM}^{-1})$	0,005	0,010	0,020	0,033	0,100
$1/\text{Verapamil bound (pmol}^{-1})$	5,00	6,06	8,33	11,11	25,00

The **KLOTZ** equation : $1/[RL] = 1/[L].(K_D/n[P_t]) + 1/n[P_t]$

So : the slope = $K_D/n[P_t]$

The y-intercept = $1/n[P_t]$



Graphically :

- y-intercept = $1/n[P_t] = 3,8 \text{ pmol}^{-1}$
- we have : $[P_t] = 0,2 \text{ mg}$ so $n = 1,31 \text{ pmol/mg}$
- The slope = $K_D/n[P_t] = 200 \text{ pmol}^{-1} \cdot \text{nM}^{-1}$ so $K_D = 52,4 \text{ nM}$

Exercise n° 5 :

We have :

- ✓ The protein : deoxyribonuclease (DNase)
- ✓ The ligand : calcium ion Ca^{2+}
- ✓ Technique used : gel filtration

The SCATCHARD graph plots $[RL]/[L]$ as a function of $[RL]$, as follows :

$$[\text{Ca}^{2+}]_{\text{bound}}/[\text{Ca}^{2+}]_{\text{free}} = f([\text{Ca}^{2+}]_{\text{bound}})$$

Note: the number of Ca^{2+} ions bound per enzyme molecule = $[\text{Ca}^{2+}]_{\text{bound}}/[P_t]$

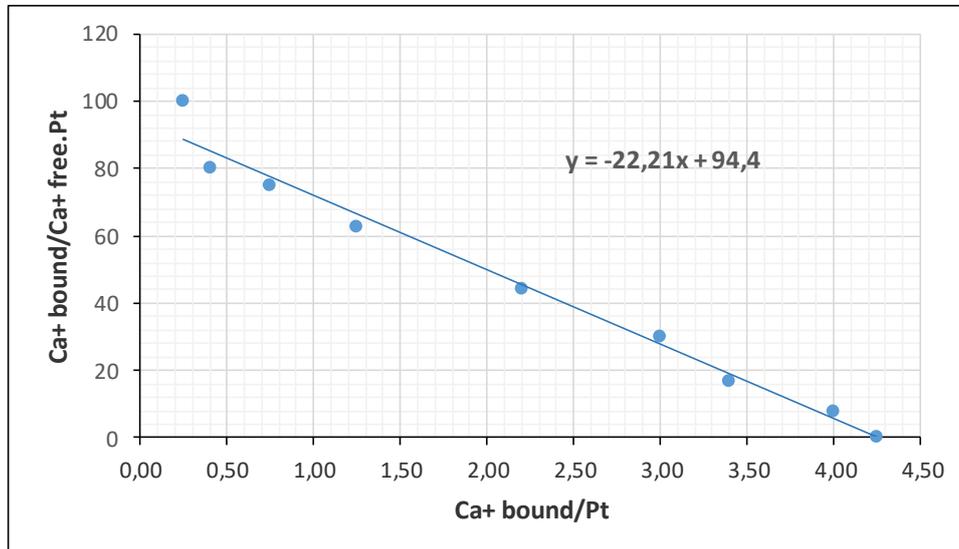
$[\text{Ca}^{2+}]_{\text{bound}}/[P_t]$	4,00	3,40	3,00	2,20	1,25	0,75	0,40	0,25
$[\text{Ca}^{2+}]_{\text{bound}}/[\text{Ca}^{2+}]_{\text{free}} \cdot [P_t]$ ($\times 10^{-3}$) (μM^{-1})	8	17	30	44	62,5	75	80	100

The SCATCHARD equation is written as : $[RL]/[L] = (-1/K_D)[RL] + n[P_t]/K_D$

Since we have $[RL]/[P_t]$, this equation becomes : $[RL]/[L][P_t] = (-1/K_D)[RL]/[P_t] + n/K_D$

From which : the slope = $-1/K_D = -K_A$

X-intercept = n



Graphically :

- X-intercept = 4,4 so **n = 4 sites**
- The slope = $-20 \cdot 10^{-3} \mu\text{M}^{-1}$ so **$K_A = 20 \cdot 10^{-3} \mu\text{M}^{-1}$**
- $K_D = 1/K_A$ so **$K_D = 50 \mu\text{M}$**